

# A Computational Framework for DNA Sequencing Microscopy

Ian T. Hoffecker<sup>a</sup>, Yunshi Yang<sup>a</sup>, Giulio Bernardinelli<sup>a</sup>, Pekka Orponen<sup>b</sup>, and Björn Högberg<sup>a,2</sup>

<sup>a</sup>Department of Medical Biochemistry and Biophysics, Karolinska Institutet, S-17177 Stockholm, Sweden; <sup>b</sup>Department of Computer Science, Aalto University, FI-00076 Aalto, Finland

This manuscript was compiled on August 16, 2019

**We describe a method whereby micro-scale spatial information such as the relative positions of biomolecules on a surface can be transferred to a sequence-based format and reconstructed into images without conventional optics. Barcoded DNA polony amplification techniques enable one to distinguish specific locations of a surface by their sequence. Image formation is based on pairwise fusion of uniquely tagged and spatially adjacent polonies. The network of polonies connected by shared borders forms a graph whose topology can be reconstructed from pairs of barcodes fused during a polony crosslinking phase, the sequences of which are determined by recovery from the surface and next-gen sequencing. We developed a mathematical and computational framework for this principle called Polony Adjacency Reconstruction for Spatial Inference and Topology and show that Euclidean spatial data may be stored and transmitted in the form of graph topology. Images are formed by transferring molecular information from a surface of interest, which we demonstrated *in silico* by reconstructing images formed from stochastic transfer of hypothetical molecular markers. The theory developed here could serve as a basis for an automated, multiplexable, and potentially super resolution imaging method based purely on molecular information.**

next gen sequencing | DNA microscopy | polonies | DNA computing | graph theory |

Microscopic imaging has traditionally relied on optics to amplify signals derived from initially confined spatial regions. Exceptions include atomic force microscopy which images by utilizing a probe to interact with the sample. DNA has a high information density, with storage levels of 5.5 petabits per cubic millimeter achieved (1), making it an attractive medium for encoding spatial information at microscales. In this paper, we present a theoretical foundation for a spatial information encoding approach that utilizes DNA sequencing and graph theory that could be used to generate whole images.

DNA-driven reactions can be coupled to optically-acquired spatial information such as with proximity ligation assay (PLA) (2), and DNA-PAINT (3) where molecular interactions mediated by DNA are discovered using fluorescence. There is also a family of techniques for connecting spatial locations with single cell RNA sequencing data: using *a priori* knowledge of spatial marker genes to associate unknown genes to approximate locations, the *a priori* data being in most cases obtained by microscopy such as with *in situ* hybridization or modelling of spatial expression patterns to retrieve locations of associated genes (4–9). Alternatively, direct microscopy-based *in situ* sequencing methods achieve precise context-sensitive spatial transcriptomic information without needing to scramble spatial data by dissociation prior to sequencing (10, 11).

Encoding spatial information in a way that is preserved in the scrambling during isolation and recovery from *in situ*

contexts that can then be read and recovered with sequencing is a major challenge. A few techniques achieve this by encoding spatial information directly into a molecular format, e.g. in the form of DNA read during sequencing along with transcriptomic data. These methods are based on artificial generation of an addressable surface using printing or lithography (12–14).

Herein, we describe a computational framework for a method called Polony Adjacency Reconstruction for Spatial Inference and Topology (PARSIFT), for the purpose of encoding images, for example of the positions of specific molecules relative to others on a 2D plane, directly into a DNA-based format without transduction of information through any other medium without *a priori* surface addressing. PARSIFT utilizes the connectivity of vertices in a graph of paired DNA sequences to infer Euclidean spatial adjacency and next-gen sequencing to recover that information *a posteriori*.

Encoding of topological data in DNA sequence format is possible by using DNA barcodes (unique molecular identifiers), i.e. randomized stretches of bases within a sequence of synthetic DNA. Barcodes associated with spatial patches can establish an identity for those locations, each patch distinguishable from another by sequence. A DNA barcode with 10 bases has over a million possible sequences, and larger barcodes can be used to create effectively unique labels in a system. The basic unit of topological data is an edge or association between two adjacent patches by physically linking between their barcodes. Topological mapping with barcoding has been used to infer neural connectomes by building a network from cells sharing common barcodes left by cell-traversing viruses (15) as well as features of DNA origami (16).

We can barcode surface patches using polony generation methods like bridge amplification (17), a 2-primer rolling

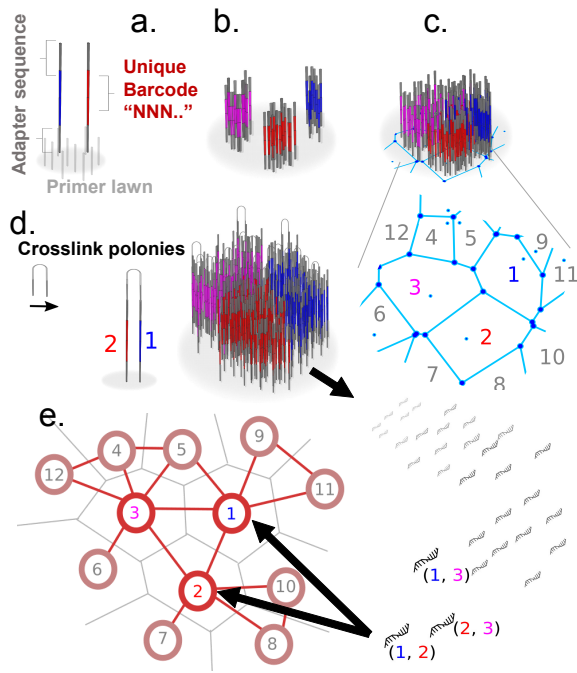
## Significance Statement

Traditional microscopy is based on the propagation of interactions between light and small scale objects up to larger scales. Such information may be encoded in DNA and transmitted with next-gen sequencing to be later reconstructed and visualized computationally. We provide a mathematical framework and computational proof of concept for a form of DNA-sequencing based microscopy that may be used to construct whole images without the use of optics. Such an approach can be automated in a parallel and multiplexable way that current optical and scanning-based techniques are unable to achieve.

ITH, GB, and BH conceived project. ITH and YY implemented the *in silico* proof of concept. ITH, YY, PO, and BH developed the mathematical theory. ITH, YY, GB, PO, and BH wrote the manuscript.

The authors declare no conflict of interest.

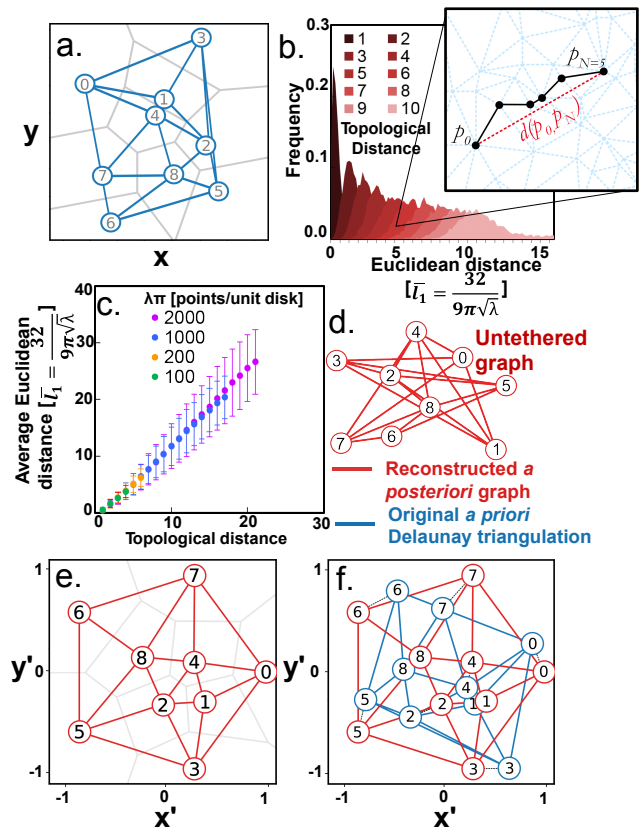
<sup>1</sup>To whom correspondence should be addressed. E-mail: bjorn.hogberg@ki.se



**Fig. 1.** Encoding and recovering metrics through polony adjacency. (a) Seed molecules with unique barcode sequences land randomly on a surface of primers. (b) Local amplification of seed molecules produces sequence-distinct polonies. (c) Saturation of polonies occurs when polonies are blocked from further growth by encountering adjacent polonies, forming a tessellated surface. (d) Random crosslinking of adjacent strands leads to pairwise association of nearby barcodes. (e) Recovery and sequencing of barcode pairs enables reconstruction of a network with similar relative positions of polonies as the original surface.

59 circle amplification (18), template walking amplification (19),  
 60 or packing of barcoded beads (20). Unique “seed” strands are  
 61 captured by primer strands on the surface (Figure 1a) and  
 62 locally amplified in the immediate vicinity where they landed.  
 63 This generates numerous distinct patches, or “polonies”, of  
 64 amplified DNA (Figure 1b). Within each, all DNA is derived  
 65 from a single seed molecule. Any of the above techniques  
 66 could be applied to our method, though we focus herein on  
 67 the polony-amplification-by-surface-primers approach.

68 By growing polonies on a surface of primers to saturation  
 69 (Figure 1c), i.e. when growing polonies encounter the bound-  
 70 aries of other adjacent polonies, a tessellation of neighboring  
 71 polonies forms. Each polony has a limited number of immedi-  
 72 ately adjacent neighboring polonies with their own respective  
 73 barcodes. Though each patch is associated with a unique  
 74 sequence according to its parent seed molecule, isolation of  
 75 this DNA and subsequent sequencing would scramble informa-  
 76 tion about the polony’s position and its neighboring polonies.  
 77 Thus the critical step is to crosslink strands (SI Appendix Fig.  
 78 S1) from each polony to strands from adjacent polonies (Fig-  
 79 ure 1d) in a way that enables both barcodes to be sequenced  
 80 together in a single read. Recovery of the strands, i.e. strip-  
 81 ping them from the surface followed by next-gen sequencing  
 82 (by any means including non-optical approaches such as Ox-  
 83 ford Nanopore) thus preserves topological association between  
 84 neighboring polonies as pairs of barcodes — a complete set of  
 85 which constitutes the whole topological network of adjacent  
 86 polonies (Figure 1e). For random seed distributions we show  
 87 that topological information alone, constrained by being a



**Fig. 2.** Encoding and recovering metrics via topology. (a) Nine seed molecule points distributed randomly on a plane, the induced Voronoi tessellation  $T$  (gray lines), its Delaunay diagram  $D$  (blue lines), and the untethered graph  $G$ . (b) The distribution of Euclidean distances associated with a given topological distance (path with the fewest edges between two points) sampled for random Poisson Delaunay triangulations (5000 samples per topological distance value). (c) Euclidean distances normalized to the average length of a typical Poisson Delaunay edge (Equation 9.9 (21)) plotted versus topological distance for different Poisson intensities, exhibiting linearity between topological and Euclidean distance. (d) The untethered graph: a set of nodes (black) and edges (red) that constitutes the information preserved after dissociation from spatial context. (e) Reconstructed planar embedding of the initially untethered graph (red lines) using the Tutte embedding approach and corresponding Voronoi tessellation (gray lines). (f) Alignment of reconstructed embedding from e with the original Delaunay diagram from a.

2D planar network with known boundary geometry, retains  
 significant spatial metrics of the original distribution. By gen-  
 erating such a mappable surface, we propose that localization  
 of molecules bound to the surface can be done by covalent  
 association with polonies, enabling inference of molecular spa-  
 tial distributions and construction of images with polonies as  
 pixels.

## 1. Results and Discussion

**A. Voronoi Tessellation as a Model of Polony Saturation.** The  
 spatial distribution of polonies on a surface, the *a priori* Eu-  
 clidean information that is not explicitly accessible after isola-  
 tion, can be preserved by associations between adjacent polony  
 sequences and recovered with sequencing. Information that is  
 available after sequencing and subsequent transformations of  
 that data are then referred to as *a posteriori*.

Assume that seed molecule amplification on a bounded 2D  
 surface, say in the shape of a disk, takes the form of uniform

105 circular growth. At the point of saturation, polonies have  
 106 amplified to the extent that their expanding boundaries are  
 107 restricted from further growth, having encountered neighboring  
 108 polonies. The system of polonies then forms a planar *Voronoi*  
 109 *tessellation*  $T$  (SI Appendix A), appearing as a characteristic  
 110 mosaic of polygons with the property that every point within  
 111 a given cell is closer to its parent seed point than any others.  
 112  $T$  can also be represented by its plane dual *Delaunay diagram*  
 113  $D = (P, L)$  whose vertices  $P$  are the seed points of  $T$  and  
 114 edges  $L$  are the line segments connecting the seed points of  
 115 adjacent cells (polonies). By the geometric characteristics of  
 116  $T$ , all the faces of  $D$  are triangles (22)(Section 9).

117 We refer to the graph defined purely by its vertices and  
 118 edges without spatial considerations as the *untethered graph*.  
 119 Figure 2a presents a miniature Voronoi tessellation  $T$  formed  
 120 from 9 seed points within a square and its Delaunay diagram  
 121  $D$ . The untethered graph  $G = (V, E)$  (Figure 2d) is obtained  
 122 from  $D$  by omitting all geometric information, retaining only  
 123 topological characteristics of the Delaunay diagram  $D$ . This  
 124 includes a topological distance function  $t(i, j)$  defined as the  
 125 fewest number of edges that must be traversed to get from  
 126 one vertex  $i$  to another  $j$ , but no other information about  
 127 the spatial origins of  $G$  is explicitly stored (e.g. no Euclidean  
 128 coordinates of the original points).

**B. Topological Metrics as a Proxy for Euclidean Metrics.** Let  
 $P = \{p_k \mid k = 1, \dots, N\}$  be a planar placement of  $N$  seed  
 points, resulting from a Poisson-distributed seeding with in-  
 tensity (i.e. polony density)  $\lambda$  over an area  $A$ . Thus  $N \approx \lambda A$ ,  
 and an untethered graph representation  $G = (V, E)$  of the  
 true Delaunay diagram  $D$  can be obtained by:

$$V = \{1, \dots, N\},$$

$$E = \{\{i, j\} \mid \text{barcodes } w_i \text{ and } w_j \text{ co-occur, } i, j = 1, \dots, N\}$$

129 Since sufficiently long barcodes are with high probability  
 130 unique (SI Appendix B), we treat pairs of barcodes as unique  
 131 markers of polony adjacency. We postulate that with a suffi-  
 132 ciently dense Poisson-distributed placement  $P$ , the topological  
 133 metric on  $G$  (with an appropriate linear scaling) approxi-  
 134 mates well the actual Euclidean metric of the points in  $P$   
 135 (SI Appendix D-E). Figure 2b shows the Euclidean distance  
 136 distributions for increasing topological distances from a refer-  
 137 ence vertex, for a large collection of Delaunay triangulations  
 138 of Poisson random point sets. Figure 2c then plots the scaled  
 139 (Equation 9.9 (21)) average Euclidean distances as a func-  
 140 tion of topological distances for Delaunay triangulations of  
 141 random point sets generated by Poisson processes of increas-  
 142 ing intensity  $\lambda$ , showing crucially that the two variables are  
 143 proportional.

144 On this basis we propose that by finding a proper straight-  
 145 line planar embedding of the untethered graph  $G$  we approxi-  
 146 mate also the metric properties of the underlying Delaunay  
 147 diagram  $D$  and the corresponding Voronoi tessellation  $T$ . A  
 148 straight-line embedding of  $G$  in a plane is determined by the  
 149 placement  $P'$  of its vertices, from which the line segments  
 150  $L'$  corresponding to the edges can be deduced, thus denoted  
 151 as  $\langle G, P' \rangle$ . Our hidden *a priori* embedding is the Delaunay  
 152 diagram  $D = \langle G, P \rangle$ , and the goal is to approximate this with  
 153 a good *a posteriori* embedding  $\langle G, P' \rangle$ .

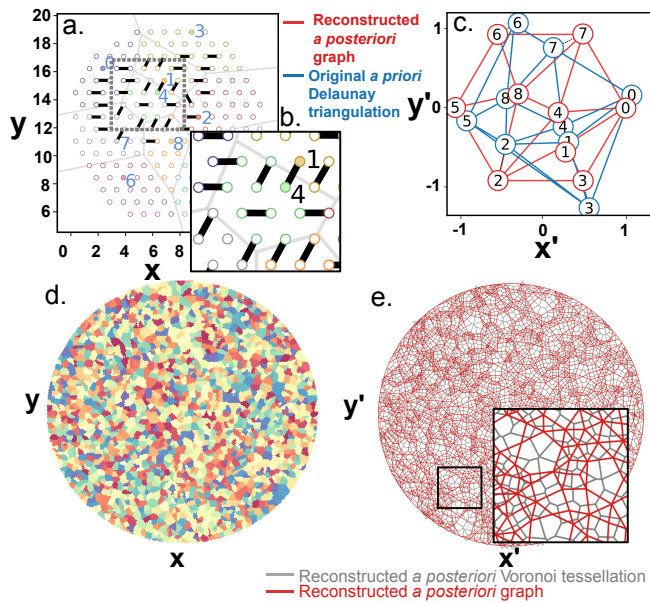
154 One constraint on our candidate  $\langle G, P' \rangle$  is that it must be  
 155 *planar*, i.e. no two edges may cross each other. This is due to

156 the physical assumption that the barcode-pairings correspond  
 157 to polony adjacencies and thus cannot bridge non-neighboring  
 158 polonies. There are several efficient algorithms for finding a  
 159 plane embedding of a planar graph, one of which is the *Tutte*  
 160 or *barycentric embedding* (23), applicable to Delaunay-diagram  
 161 type graphs. Another quality constraint is that an average  
 162 spatial density of the *a posteriori* vertex positions  $\lambda'$  should  
 163 be obtained from the final distribution with no systematic  
 164 variation across the reconstructed area. Finally, if we were to  
 165 generate a new Delaunay triangulation from the reconstructed  
 166 points (as can be done from any arbitrary set of points), this  
 167 should produce a similar set of edges as the original untethered  
 168 graph that was the basis for reconstruction.

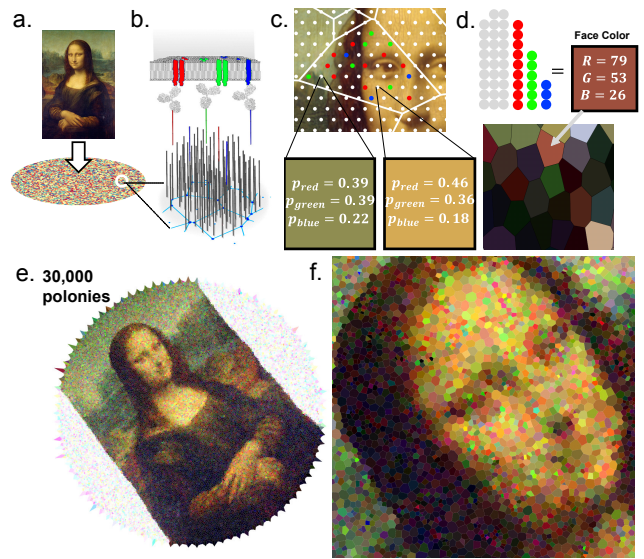
169 Our reconstruction approach (flow diagram SI Appendix  
 170 Fig. S2) starts by determining the *outer* or *boundary face* of  
 171 the Delaunay diagram  $D$  underlying the untethered graph  $G$ .  
 172 This can practically be done by finding the face in any planar  
 173 embedding of  $G$  that has the most vertices with an intermediate  
 174 planar embedding, because in  $D$  all faces except the boundary  
 175 face are few-vertex triangles. Fixing the placement of the  
 176 vertices on the boundary face, we then compute positions for  
 177 the other vertices of  $G$  by Tutte's algorithm, which simply  
 178 places each vertex at the average (barycenter) of its neighbors'  
 179 positions. In the case of a Delaunay-diagram type graph with  
 180 the boundary face a convex polygon, this system is guaranteed  
 181 to be non-degenerate (23), and the result will be a crossing-free  
 182 straight-line embedding of  $G$ .

183 If spatial characteristics of the original Euclidean boundary  
 184 are known — for instance if we specify that all boundary points  
 185 must lie on a circle of known radius — then the embedding  
 186 may also be scaled to match the original Euclidean metrics.  
 187 Figure 2e shows the Tutte embedding of the untethered graph  
 188 (Figure 2d) with boundary points arranged uniformly around  
 189 the unit disk. For comparison, we have aligned the recon-  
 190 structed graph with the original Delaunay diagram (Figure 2f)  
 191 by linearly transforming the planar graph to minimize the  
 192 distance between corresponding vertices. We can see that  
 193 relative positions are preserved albeit with local distortion  
 194 that leads to slight displacement of each reconstructed vertex  
 195 relative to its original seed counterpart. The algorithm thus  
 196 returns approximate relative spatial positions of polonies from  
 197 an input of paired polonies.

**C. Simulation and Reconstruction by Embedding.** We simu-  
 198 late the primer lawn as a hexagonally packed disk of area  
 199  $A$  with  $M$  primer sites as the region of interest (ROI) (Fig-  
 200 ure 3a). We simulate a random seeding at a polony density  
 201  $\lambda$  by selecting  $N = \lambda A$  random sites, followed by pairing of  
 202 adjacent polony primer sites and scrambling of edge data prior  
 203 to reconstruction. Figure 3b shows how crosslinking leads  
 204 to random pairing of adjacent sites, some of which are self-  
 205 pairing events (providing no additional pairing information)  
 206 and some of which are cross-polony sites that can be used to  
 207 deduce the presence of a spatial boundary, with the fraction  
 208 of information-bearing cross-pairs diminishing with the rela-  
 209 tive site density  $\rho \stackrel{\text{def}}{=} M/(A\lambda)$  or average number of sites per  
 210 polony (SI Appendix Fig. S3). The probabilistic nature of the  
 211 pairing opens up the possibility to miss an existing boundary,  
 212 particularly when the boundary is small or when  $\rho$  is low. A  
 213 2000 polony-simulated surface is shown in Figure 3c, and SI  
 214 Appendix Fig. S4 shows site-linking and the corresponding  
 215



**Fig. 3.** Simulation of polony adjacency reconstruction. (a) Lattice diagram of primer lawn and polonies denoted with color and Voronoi cell boundaries. Filled circles indicate seed locations. (b) Illustration of random site pairing between adjacent primer sites. (c) Alignment between *a priori* and *a posteriori* points from a. (d) Larger simulated surface with a polony density  $\lambda = 2000$  polonies/unit area and a relative site density  $\rho = 50$  sites per polony on average. (e) Reconstructed graph (red lines) and corresponding Voronoi tessellation (gray lines) computed using the Tutte embedding approach from scrambled edges derived from the simulated surface in d.



**Fig. 4.** Voronoi image formation. (a) An image is overlaid on a surface of primer sites. (b) Molecular markers representing different targets (R, G, and B) contact-transferred to the polony surface and each covalently linked to a polony barcode. (c) Monte Carlo sampling to determine if a marker is associated with a given site and if so which target by taking the probability from the RGB value normalized to 1 at the corresponding position in the image. (d) Tallying of markers and empty sites within a polony/Voronoi cell determines the color and brightness of that “pixel”. A subsequent image (lower pane) is formed by coloring each cell accordingly. (e) Larger scale reconstruction from scrambled edge data using the Tutte embedding approach with 30,000 polonies. (f) Closeup of e revealing individual Voronoi “pixels”

216 Delaunay triangulation of a 500 polony example.

217 We reconstructed the topological network from the scrambled edges and performed intermediate embedding, boundary  
 218 face determination, and Tutte embedding (Figure 3c). For  
 219 this larger reconstruction, spatial uniformity is more appar-  
 220 ent, we see that Voronoi cells take on the approximate size of  
 221 polonies in the *a priori* surface, observe no obvious systematic  
 222 changes in mesh density across the length of the ROI, and note  
 223 the absence of crossed edges. Besides the Tutte embedding  
 224 strategy, we developed 2 additional approaches for approxi-  
 225 mating Euclidean metrics from the untethered graph. One  
 226 is a non-deterministic spring relaxation (24). This approach  
 227 does not strictly require a crossing-free planar embedding, and  
 228 can thus lead to provably false positions involving non-planar  
 229 adjacency, however this feature could also be advantageous  
 230 if natural interpenetration of adjacent polonies leads to such  
 231 topology. The last approach (SI Appendix F) is based on the  
 232 notion of topological distance  $t(i, j)$  and its role as a proxy  
 233 for Euclidean distance. We extend the principle of geometric  
 234 triangulation, whereby the set of distances of a point to other  
 235 points in a plane can be converted to Cartesian coordinates, to  
 236 incorporate  $t(i, j)$  as a surrogate for Euclidean distance. In one  
 237 variant of this method, a total topological distance matrix is  
 238 reduced to two principal component vectors approximating the  
 239  $x$  and  $y$  coordinate vectors. In the alternative variant,  $t(i, j)$   
 240 of each vertex are only measured out to peripheral vertices, re-  
 241 ducing systematic distortions. A comparison of reconstructed  
 242 meshes from the different approaches is shown in SI Appendix  
 243 Fig. S5-S6.

245 **D. Stamping and Image Formation.** Knowledge of polony loca-  
 246 tions could be exploited to provide spatial information about

247 objects of interest. We devised a basic model of image re-  
 248 construction from the principle of contact or diffusion-based  
 249 transfer of molecules of interest to the mapped surface, i.e.  
 250 a kind of molecular stamp. As proof of concept, we use an  
 251 image (Figure 4a) as a representation of a hypothetical proba-  
 252 bility distribution of 3 types of molecular markers labeled with  
 253 identifying sequences called “red”, “green”, and “blue”. The  
 254 image represents a surface of interest that we would like to  
 255 sample from, for example a cell surface covered in oligo-tagged  
 256 antibodies, each of which would be coupled enzymatically to  
 257 a given polony upon contact (Figure 4b) or diffusing RNA  
 258 molecules like in (14). The color of the image corresponds to  
 259 the density of such markers and thus the probability that a  
 260 marker of a particular color is placed on the polony surface.  
 261 To simulate molecule transfer, the overlaid lattice of primer  
 262 sites denotes points where a Monte Carlo sampling will occur  
 263 in the corresponding position in the image. If the image pixel  
 264 at a given primer site location has an RGB value dominated by  
 265 red and green for example, then there is a higher probability  
 266 of that site being occupied by either a green or red marker  
 267 (Figure 4c). Realistically, molecular transfer introduces distor-  
 268 tion, e.g. from curvature of cell membranes or lateral diffusion  
 269 of mRNAs.

270 According to the reconstruction procedure, a Voronoi tes-  
 271 sellation is produced from the final set of vertex positions -  
 272 each cell of which constitutes a pixel that can be used to form  
 273 an image. The final RGB value of the cell can be determined  
 274 by tallying the markers that have associated with the primer  
 275 sites in the polony as well as the number of un-associated sites  
 276 (Figure 4d). The Voronoi-images shown in Figure 4e and f  
 277 were generated with the scrambling step that removes any

278 spatial information of the original image and reconstructed  
 279 using our algorithm. Note that global rotation and chirality  
 280 are not explicitly preserved from the original image. To place  
 281 this 30,000 pixel image in experimentally relevant terms, we  
 282 point to a recent spatial transcriptomics manuscript (20)  
 283 where circular discs of barcoded 10  $\mu\text{m}$  beads (in their case  
 284 sequenced optically *in situ* to obtain sequence addresses) are  
 285 used to capture transcriptomic data from tissue slices. Ro-  
 286 driques *et al* report a typical size of 70,000 10  $\mu\text{m}$  beads per  
 287 3 mm disk and obtain approximate single-cell resolution (see  
 288 also SI Appendix H). Image reconstructions from the four  
 289 approximation approaches are compared in SI Appendix Fig.  
 290 S5-S6.

291 **E. Assessment of Distortion and Precision.** We may charac-  
 292 terize reconstruction quality by defining a distortion metric.  
 293 The *a priori* seed distribution points have a 1-to-1 corre-  
 294 spondence with points in the *a posteriori* reconstruction, and  
 295 since we generated the *a priori* points ourselves, we can di-  
 296 rectly compare corresponding original and inferred positions  
 297 by applying a linear transform  $Tx(P)$  (rotation, mirroring,  
 298 scaling, and translation) to the set of reconstructed points  
 299 that minimizes net displacement between the two distributions.  
 300 Distortion is thus defined as the set of displacements:  $Df =$   
 301  $(Df_1, \dots, Df_N) \stackrel{\text{def}}{=} d(P, Tx(P')) \mid \min(\sum_{i=1}^N d(p_i, Tx(p'_i)))$ . Av-  
 302 eraged over multiple runs, we obtain 2D histograms (Figure 5a  
 303 and SI Appendix Fig. S7-S8) of distortion as a function of  
 304 position in the ROI. Increasing the polony density ( $\lambda$ ) re-  
 305 duces average distortion  $\overline{Df} = \frac{1}{N} \sum_{i=1}^N Df_i$  (Figure 5d and SI  
 306 Appendix Fig. S10) whereas changes in the site density  $\rho$  (Fig-  
 307 ure 5f and SI Appendix Fig. S11) has a negligible effect on  $\overline{Df}$   
 308 except at  $\rho < 100$  sites per polony near the point of network  
 309 disconnection from absent edges. Examining a single simula-  
 310 tion (Figure 5b) we can visualize typical distortions, persistent  
 311 over limited local scales and occurring with greater probability  
 312 near the boundaries. Analysis of the radial distribution of  
 313 this instance (Figure 5c) reveals this as a mild systematic  
 314 worsening near the boundary, an artifact introduced by the al-  
 315 gorithm's treatment of vertices on the boundary. SI Appendix  
 316 Fig. S9 compares single instance distortions for the different  
 317 reconstruction approaches.

318 We also characterize reconstruction quality with Leven-  
 319 shtein distance ( $lev_{G,G'}$ ), the number of edits needed to make  
 320 two graphs identical, between the untethered graph and set  
 321 of edges derived from a Delaunay triangulation  $D'$  generated  
 322 from the final reconstructed coordinates. Importantly, this  
 323 metric is based only on *a posteriori* information, so it can  
 324 be used in an experimental context where knowledge of the  
 325 underlying distribution is unavailable. It weakly but positively  
 326 correlates with distortion for a given  $\lambda$  (SI Appendix Fig. S13).  
 327  $lev_{G,G'}$  grows linearly with  $\lambda$  (Figure 5e and SI Appendix Fig.  
 328 S10), and like distortion is relatively constant as a function  
 329 of  $\rho$  with a transient catastrophic breakdown at low  $\rho$  (Fig-  
 330 ure 5g SI Appendix Fig. S11). We also measured a classical  
 331 resolution, the full width half maximum (FWHM) of a point  
 332 spread function (Figure 5h), by sampling the inferred posi-  
 333 tion of a single site (taking its position to be the centroid of  
 334 whatever Voronoi cell it lands in). Like distortion, FWHM is  
 335 approximately  $\propto 1/\sqrt{\lambda}$  (Figure 5i) indicating that to halve the  
 336 minimum size of distinguishable features, one should quadru-  
 337 ple  $\lambda$  (SI Appendix Fig. S12). In experimental terms, polonies

338 generated from techniques like template walking amplification,  
 339 which forms polonies from sites that must be near the packing  
 340 limit of oligo surface immobilization, can be on the order of  
 341 nanometers (19) (SI Appendix G).

## 2. Discussion and Conclusion 342

343 The three reconstruction methods (Tutte embedding, spring  
 344 relaxation, and topological distance matrix) succeed in pro-  
 345 ducing approximations of the original seed distributions that  
 346 can be used to generate images. Tutte embedding exhibited  
 347 the best estimated algorithmic complexity (based on run time  
 348 scaling with  $\lambda$ , SI Appendix Fig. S14) making it the fastest  
 349 technique which becomes significant for large reconstruction  
 350 problems ( $\lambda > 10,000$  polonies/unit area). Both Tutte embed-  
 351 ding and spring relaxation had the lowest distortion levels, with  
 352 Tutte embedding exhibiting slightly better  $Df$  and  $lev_{G,G'}$   
 353 scaling with  $\lambda$ . Tutte embedding was sensitive to catastrophic  
 354 failure at low  $\rho$ , with singly-connected edges crashing the  
 355 reconstruction, and all four approaches were sensitive to dis-  
 356 joint subgraphs - making noisy and unconnected graph data  
 357 a likely challenge for experimental scenarios. SI Appendix  
 358 Fig. S13 and SI Appendix I discuss our attempts to move  
 359 towards an algorithm that optimally exploits the available  
 360 information, and future research should seek to establish a  
 361 provably maximum-entropy reconstruction that is efficient and  
 362 deterministic.

363 Along these lines, utilizing information such as the number  
 364 of self-pairing events could be useful to extract more informa-  
 365 tion and weight edges according to estimated polony size and  
 366 better control point placement. Alternatively, low-information  
 367 content self-pairing events could be prohibited through a bipar-  
 368 tite network approach whereby only pairings between A-type  
 369 and B-type polonies would be allowed (SI Appendix Fig. S15).  
 370 The bridge amplification approach to polony generation leaves  
 371 the possibility of doing this with two species of independent  
 372 primers on the surface and two interpenetrating/overlapping  
 373 and independently saturated polony surfaces. Another possi-  
 374 ble approach is series growth of polonies. In the basic concept  
 375 presented in previous sections, a primer of uniform sequence is  
 376 assumed, however generation of a saturated layer of polonies  
 377 that could then be used as primers for a subsequent polony  
 378 generation step would then result in an overlapping of every  
 379 2nd-layer polony with multiple 1st-layer polonies. This would  
 380 result in efficient pairing of barcodes without the need for  
 381 subsequent crosslinking steps.

382 At the time of publication, we are aware of immediately  
 383 prior works whose contributions are complementary to ours  
 384 on development of DNA-sequencing based microscopy (25,  
 385 26). The former work experimentally demonstrates DNA  
 386 microscopy with images of mRNA in cells using locally confined  
 387 cDNA amplifications and polymerase extension-based fusion  
 388 of barcodes to connect spatial patches. Their approach differs  
 389 from ours through the fact that fusion events are used as  
 390 a direct distance metric, whereas our data instead relies on  
 391 topology as a proxy for Euclidean metrics. The latter work uses  
 392 series proximity ligation to associate planar spatial patches  
 393 and form a network, utilizing a spring relaxation approach for  
 394 reconstruction.

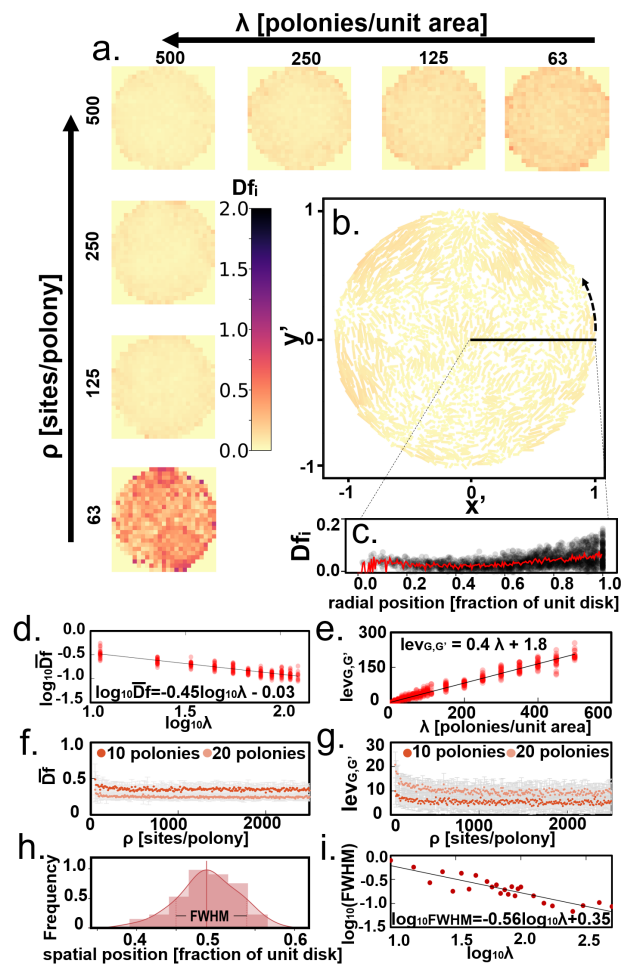
395 **A. Conclusion.** PARSIFT is a concept for microscopic image  
 396 reconstruction using spatial information encoding in DNA base

397 format. We showed an *in silico* proof of concept by construct-  
 398 ing a pipeline for taking decoupled edge data, generated from  
 399 simulated polony distributions, that are then reassembled into  
 400 a topological network and embedded in a Euclidean plane, re-  
 401 suming spatial characteristics of the original seed distribution.  
 402 We saw that global distortions are low enough to resolve whole  
 403 images. We hold that this framework and pipeline for recon-  
 404 struction could be exploited for image acquisition of micro-  
 405 and nano-scale surfaces with molecular libraries of potentially  
 406 very high multiplicity and with throughput automated in a  
 407 way that would not be possible with most optical approaches.

408 **Supporting Information (SI).** The code is available at  
 409 <https://github.com/Intertangler/parsift>

410 **ACKNOWLEDGMENTS.** The authors thank Ferenc Fördös for  
 411 insightful discussions. This work was supported by Åke Wiberg  
 412 Stiftelsen grant for medical research M17-0214 to ITH., the Knut  
 413 and Alice Wallenberg project grant 2017.0114, the Knut and Alice  
 414 Wallenberg Academy Fellow grant KAW2014.0241 to BH, and by  
 415 Academy of Finland grant 311639 to PO.

- 416 1. Church GM, Gao Y, Kosuri S (2012) Next-generation digital information storage in DNA. *Science* p. 1226355.
- 417 2. Söderberg O, et al. (2006) Direct observation of individual endogenous protein complexes in situ by proximity ligation. *Nature Methods* 3(12):995.
- 418 3. Jungmann R, et al. (2010) Single-molecule kinetics and super-resolution microscopy by fluorescence imaging of transient binding on DNA origami. *Nano Letters* 10(11):4756–4761.
- 419 4. Wang G, Moffitt JR, Zhuang X (2018) Multiplexed imaging of high-density libraries of RNAs with MERFISH and expansion microscopy. *Scientific Reports* 8(1):4847.
- 420 5. Karaiskos N, et al. (2017) The *Drosophila* embryo at single-cell transcriptome resolution. *Science* 358(6360):194–199.
- 421 6. Satija R, Farrell JA, Gennert D, Schier AF, Regev A (2015) Spatial reconstruction of single-cell gene expression data. *Nature Biotechnology* 33(5):495.
- 422 7. Achim K, et al. (2015) High-throughput spatial mapping of single-cell RNA-seq data to tissue of origin. *Nature Biotechnology* 33(5):503.
- 423 8. Halpern KB, et al. (2017) Single-cell spatial reconstruction reveals global division of labour in the mammalian liver. *Nature* 542(7641):352.
- 424 9. Lein E, Borm LE, Linnarsson S (2017) The promise of spatial transcriptomics for neuroscience in the era of molecular cell typing. *Science* 358(6359):64–69.
- 425 10. Wang X, et al. (2018) Three-dimensional intact-tissue sequencing of single-cell transcriptional states. *Science* p. eaat5691.
- 426 11. Ke R, et al. (2013) In situ sequencing for rna analysis in preserved tissue and cells. *Nature Methods* 10(9):857.
- 427 12. Lee JH, et al. (2015) Fluorescent in situ sequencing (fisseq) of rna for gene expression profiling in intact cells and tissues. *Nature Protocols* 10(3):442.
- 428 13. Crosetto N, Bienko M, Van Oudenaarden A (2015) Spatially resolved transcriptomics and beyond. *Nature Reviews Genetics* 16(1):57.
- 429 14. Ståhl PL, et al. (2016) Visualization and analysis of gene expression in tissue sections by spatial transcriptomics. *Science* 353(6294):78–82.
- 430 15. Peikon ID, et al. (2017) Using high-throughput barcode sequencing to efficiently map connectomes. *Nucleic Acids Research* 45(12):e115–e115.
- 431 16. Schaus TE, Woo S, Xuan F, Chen X, Yin P (2017) A DNA nanoscope via auto-cycling proximity recording. *Nature Communications* 8(1):696.
- 432 17. Adessi C, et al. (2000) Solid phase DNA amplification: characterisation of primer attachment and amplification mechanisms. *Nucleic Acids Research* 28(20):e87–e87.
- 433 18. Korfhage C, et al. (2017) Clonal rolling circle amplification for on-chip DNA cluster generation. *Biology Methods and Protocols* 2(1).
- 434 19. Ma Z, et al. (2013) Isothermal amplification method for next-generation sequencing. *Proceedings of the National Academy of Sciences* 110(35):14320–14323.
- 435 20. Rodrigues SG, et al. (2019) Slide-seq: A scalable technology for measuring genome-wide expression at high spatial resolution. *Science* 363(6434):1463–1467.
- 436 21. Miles RE (1970) On the homogeneous planar Poisson point process. *Mathematical Biosciences* 6:85–127.
- 437 22. de Berg M, van Krefeld M, Overmars M, Cheong O (2008) *Computational Geometry: Algorithms and Applications, 3rd Rev. Ed.* (Springer-Verlag).
- 438 23. Tutte WT (1963) How to draw a graph. *Proceedings of the London Mathematical Society* 3(1):743–767.
- 439 24. Kamada T, Kawai S, et al. (1989) An algorithm for drawing general undirected graphs. *Information Processing Letters* 31(1):7–15.
- 440 25. Weinstein JA, Regev A, Zhang F (2019) DNA microscopy: Optics-free spatio-genetic imaging by a stand-alone chemical reaction. *Cell*.
- 441 26. Boulgakov A, Xiong E, Bhadra S, Ellington AD, Marcotte EM (2018) From space to sequence and back again: Iterative DNA proximity ligation and its applications to DNA-based imaging. *bioRxiv*.



**Fig. 5.** Reconstruction quality. (a) 2D histograms of average displacement values binned by relative position in the unit disk ( $n = 5000/\lambda$  simulations per histogram) for varied parameters ( $\lambda$  and  $\rho$ ). (b) Distortion in a single 2000 polony Tutte embedding with lines connecting *a priori* and *posteriori* vertex locations. Color map indicates line length (max = unit disk diameter 2.0). (c) Radial profile of distortion in b and 5 point moving average (red line). (d) Log-log plot of average displacement versus  $\lambda$  (points single individual simulations reconstructions) and fixed  $\rho = 500$  sites per polony showing displacement approximately  $\propto 1/\sqrt{\lambda}$ . (e) Linear plot of Levenshtein distance ( $lev_{G,G'}$ ) between untethered and *a posteriori* Delaunay graphs as function of polony. (d and e:  $n = 25$  simulations per  $\lambda$  value) (f) Plot of average displacement and (g) plot of  $lev_{G,G'}$  each as a function of  $\rho$  for two values of  $\lambda$ , error bars represent standard deviation (f and g:  $n = 25$  simulations per point, error bars: standard dev.). (h) Single instance of full width half maximum (FWHM) of a *posteriori* point spread function of a single site. (i) Log-log plot of FWHM versus  $\lambda$ , scaling approximately according to the negative square root of polony density.